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Abstract

Sequences Analyzed

Results & Conclusions

Formylglycinamide ribonucleotide amidotransferase (FGARAT) catalyzes the fourth step in the purine biosynthetic pathway. Two forms of this enzyme have been described in the literature, a monomeric form present in eukaryotes, beta and gamma proteobacteria, and a heterotetrameric form found in most other Bacteria and Archaea. The heterotetrameric enzyme is composed of PurS, PurL and PurQ subunits in a ratio of 2:1:1, while the monomeric enzyme has a domain organization corresponding to a gene fusion of *purS-purS-purL-purQ*. To investigate the evolution of this enzyme, FGARAT sequences were retrieved from the completed genomes of different families of organisms and compared.

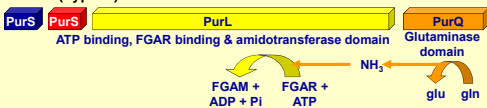
Multiple sequence alignments have revealed a heterodimeric form of the enzyme as well as a second monomeric form. The heterodimer consists of a PurQ subunit together with a PurL of intermediate length that corresponded to a *purS-purS-purL* gene fusion. The phylogenetic distribution of the heterodimer suggests that it arose in an ancestor of the delta-proteobacteria and has spread to several other organisms via horizontal gene transfer. The second monomeric FGARAT has a domain organization similar to the previously described form, but also has sequence features that suggest an independent origin. This enzyme form is present in several phylogenetically diverse organisms, thus obscuring its origin. Further insights into the evolution of FGARAT from a tetramer to a dimer and finally to a monomer have been derived from mapping of residues conserved between the different forms onto the recently solved structure of the monomeric FGARAT (Anand et al., 2004).

Organism	Taxonomy	Sub-units	PurL	PurQ	PurS
<i>Ornella foresti</i> KT0993	Bacteroidetes, Flavobacteriales	1	1225	-	-
<i>Cytophaga hutchinsonii</i>	Bacteroidetes, Spirochaetales	1	1231	-	-
<i>Saccharomyces cerevisiae</i>	Eukarya, Fungi	1	1358	-	-
<i>Homo sapiens</i>	Eukarya, Metazoa	1	1338	-	-
<i>Dictpyosium discoideum</i>	Eukarya, Mycetozoa	1	1255	-	-
<i>Arabidopsis thaliana</i>	Eukarya, Viridiplantae	1	1387	-	-
<i>Rhodoferrax ferredoxicans</i>	Proteobacteria, Beta, Burkholderiales	1	1409	-	-
<i>Polaronomas naphthalenivorans</i>	Proteobacteria, Beta, Burkholderiales	1	1340	-	-
<i>Burkholderia pseudomallei</i>	Proteobacteria, Beta, Burkholderiales	1	1356	-	-
<i>Thiosacillus denitrificans</i>	Proteobacteria, Beta, Hydrogenisporales	1	1291	-	-
<i>Methylobacillus flagellatus</i>	Proteobacteria, Beta, Methylophilales	1	1297	-	-
<i>Neisseria meningitidis</i>	Proteobacteria, Beta, Neisseriales	1	1320	-	-
<i>Nitrosomonas europaea</i>	Proteobacteria, Beta, Nitrosomonadales	1	1304	-	-
<i>Francisella tularensis</i>	Proteobacteria, Beta, Rhodocyclales	1	1210	-	-
<i>Desulfotalea psychrophila</i>	Proteobacteria, Delta, Desulfobacterales	1	1287	-	-
<i>Mycoccus xanthus</i>	Proteobacteria, Delta, Myxococcales	1	1302	-	-
<i>Aeromonas hydrophila</i>	Proteobacteria, Gamma, Aeromonadales	1	1357	-	-
<i>Colwellia psychrythraea</i>	Proteobacteria, Gamma, Alteromonadales	1	1323	-	-
<i>Baumannia cicadellitorica</i>	Proteobacteria, Gamma, Candidatus Baumannia	1	1297	-	-
<i>Nitrosococcus oceanii</i>	Proteobacteria, Gamma, Chromatiales	1	1300	-	-
<i>Salmonella typhimurium</i>	Proteobacteria, Gamma, Enterobacteriales	1	1295	-	-
<i>Escherichia coli</i>	Proteobacteria, Gamma, Enterobacteriales	1	1295	-	-
<i>Coxsilla burnii</i>	Proteobacteria, Gamma, Legionellales	1	1296	-	-
<i>Methylococcus capsulatus</i>	Proteobacteria, Gamma, Methylococcales	1	1288	-	-
<i>Nahala chejuensis</i>	Proteobacteria, Gamma, Oceanospirillales	1	1298	-	-
<i>Haemophilus influenzae</i>	Proteobacteria, Gamma, Pasteurellales	1	1320	-	-
<i>Pseudomonas syringae</i>	Proteobacteria, Gamma, Pseudomonadales	1	1313	-	-
<i>Francisella tularensis</i>	Proteobacteria, Gamma, Thiosphaerales	1	1290	-	-
<i>Vibrio fischeri</i>	Proteobacteria, Gamma, Vibrionales	1	1303	-	-
<i>Xanthomonas campestris</i>	Proteobacteria, Gamma, Xanthomonadales	1	1348	-	-
<i>Magnetococcus sp. MC-1</i>	Proteobacteria, Magnetococcales	1	1295	-	-
<i>Clostridium acetobutylicum</i>	Firmicutes, Clostridia	1	1295	-	-
<i>Clostridium perfringens</i>	Firmicutes, Clostridia	1	1298	-	-
<i>Clostridium tetani</i>	Firmicutes, Clostridia	1	1258	-	-
<i>Streptococcus pyogenes</i>	Firmicutes, Lactobacillales	1	1287	-	-
<i>Streptococcus agalactiae</i>	Firmicutes, Lactobacillales	1	1203	-	-
<i>Fusobacterium nucleatum</i>	Fusobacteria, Fusobacteriales	1	1249	-	-
<i>Corynebacterium diptheriae</i>	Actinobacteria, Actinomycetales	1	1238	-	-
<i>Bifidobacterium longum</i>	Actinobacteria, Bifidobacteriales	1	1244	-	-
<i>Methanococcus marisnigri</i>	Archaea, Euryarchaeota, Methanococcales	1	1231	-	-
<i>Methanococcus marisnigri</i>	Archaea, Euryarchaeota, Methanococcales	2	869	272	-
<i>Methanospirillum hungatei</i>	Archaea, Euryarchaeota, Methanospirillales	2	979	260	-
<i>Dehalobacterium ethanogenes</i>	Chloroflexi, Dehalococcoides	2	963	355	-
<i>Rhodospirillum rubrum</i>	Planctomycetes, Planctomycetales	2	1099	292	-
<i>Anaplasma marginale</i>	Proteobacteria, Alpha, Rickettsiales	2	1016	269	-
<i>Radiobacter bacteriovorus</i>	Proteobacteria, Delta, Bacteroidales	2	1099	293	-
<i>Desulfotalea psychrophila</i>	Proteobacteria, Delta, Desulfobacterales	2	1090	260	-
<i>Desulfobacter autotrophicus</i>	Proteobacteria, Delta, Desulfomonadales	2	996	275	-
<i>Syntrophobacter fumaroxidans</i>	Proteobacteria, Delta, Syntrophobacterales	2	1099	269	-
<i>Acidobacterium Elin345</i>	Acidobacteria, Acidobacteriales	4	768	231	80
<i>Solibacter salinarum</i>	Acidobacteria, Solibacteriales	4	742	232	81
<i>Corynebacterium glutamicum</i>	Actinobacteria, Actinomycetales	4	762	233	81
<i>Corynebacterium jeikeium</i>	Actinobacteria, Actinomycetales	4	839	223	84
<i>Streptomyces coelicolor</i>	Actinobacteria, Actinomycetales	4	782	226	80
<i>Acidothermus cellulolyticus 11B</i>	Actinobacteria, Actinomycetales	4	784	225	81
<i>Rubrobacter xylanophilus</i>	Actinobacteria, Rubrobacteriales	4	727	220	74
<i>Aquifex aeolicus</i>	Aquificae, Aquificales	4	745	227	77
<i>Pyrobaculum aerophilum</i>	Archaea, Crenarchaeota, Thermoprotei	4	697	212	84
<i>Archaeoglobus fulgidus</i>	Archaea, Euryarchaeota, Archaeoglobi	4	765	211	80
<i>Halorubrum marisnigri</i>	Archaea, Euryarchaeota, Halobacteriales	4	760	228	84
<i>Methanohalobium thermohalophilus</i>	Archaea, Euryarchaeota, Methanohalobiales	4	714	214	84
<i>Methanocaldococcus jannaschii</i>	Archaea, Euryarchaeota, Methanococcales	4	733	230	83
<i>Methanosarcina acetivorans</i>	Archaea, Euryarchaeota, Methanococcales	4	716	232	88
<i>Methanopyrus kandleri</i>	Archaea, Euryarchaeota, Methanopyri	4	724	226	84
<i>Methanococcus burtonii</i>	Archaea, Euryarchaeota, Methanococcales	4	715	231	82
<i>Pyrococcus abyssi</i>	Archaea, Euryarchaeota, Thermococci	4	705	233	84
<i>Picrophilus torridus</i>	Archaea, Euryarchaeota, Thermoplasmatia	4	741	248	75
<i>Thermoplasma acidophilum</i>	Archaea, Euryarchaeota, Thermoplasmatia	4	789	257	79
<i>Haloquadratum walsbyi</i>	Archaea, Halobacteriales, Halobacteriales	4	725	228	81
<i>Salinibacter ruber</i>	Bacteroidetes, Spirochaetales	4	754	230	82
<i>Chlorobium lipudium</i>	Chloroflexi, Chloroflexiales	4	789	234	84
<i>Synechococcus elongatus</i>	Cyanobacteria	4	777	221	74
<i>Gloeobacter violaceus</i>	Cyanobacteria, Gloeobacteriales	4	774	232	88
<i>Nostoc sp. PCC 7120</i>	Cyanobacteria, Nostocales	4	782	224	82
<i>Trichostema oyarum</i>	Cyanobacteria, Oscillatoriales	4	775	231	87
<i>Prochlorococcus marinus</i>	Cyanobacteria, Prochlorales	4	883	217	90*
<i>Deinococcus radiodurans</i>	Deinococcus-Thermus, Deinococcales	4	747	266	84
<i>Thermus thermophilus</i>	Deinococcus-Thermus, Thermales	4	725	227	84
<i>Bacillus subtilis</i>	Firmicutes, Bacillales	4	742	227	84
<i>Carboxydobacterium hydrogeniformans</i>	Firmicutes, Clostridia	4	728	234	81
<i>Thermoanaerobacter tengcongensis</i>	Firmicutes, Clostridia	4	733	224	82
<i>Moorella thermoacetica</i>	Firmicutes, Clostridia, Thermoanaerobacterales	4	733	236	84
<i>Enterococcus faecalis</i>	Firmicutes, Lactobacillales	4	739	224	83
<i>Caulobacter crescentus</i>	Proteobacteria, Alpha, Caulobacteriales	4	739	220	79
<i>Sinorhizobium meliloti</i>	Proteobacteria, Alpha, Rhizobiales	4	743	233	80
<i>Silicibacter pomeroyi</i>	Proteobacteria, Alpha, Rhizobacteriales	4	719	222	76
<i>Glucobacter oxydans</i>	Proteobacteria, Alpha, Rhodospirillales	4	734	233	80
<i>Candidatus Pelagibacter ubique</i>	Proteobacteria, Alpha, Rickettsiales	4	730	227	80
<i>Zymomonas mobilis</i>	Proteobacteria, Alpha, Spirochaetales	4	734	221	77
<i>Rhizobium etli</i>	Proteobacteria, Alpha, Rhizobiales	4	743	233	80
<i>Anaeromyxobacter dehalogenans</i>	Proteobacteria, Delta, Myxococcales	4	789	218	81
<i>Campylobacter jejuni</i>	Proteobacteria, Epsilon, Campylobacteriales	4	728	215	81
<i>Leptospira interrogans</i>	Spirochetes, Spirochaetales	4	746	219	82
<i>Thermotoga maritima</i>	Thermotogae, Thermotogales	4	683	215	82

Background

Three forms of FGARAT

- Heterotetramer (Type 4) – found in most Archaea and most Bacteria



- Heterodimer (Type 2) – found primarily in Planctomycetes and δ - proteobacteria

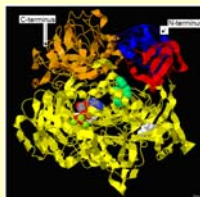


- Monomer (Type 1) – found primarily in Eukaryotes, β and γ - proteobacteria



Figure 1. Domain/ Subunit organization of FGAR Amidotransferase Enzymes.

Color coding of subunits and domains correspond to domain colors shown in 3-D structure view of monomeric enzyme found in *Salmonella typhimurium* (Anand et al., 2004).



- Goals: 1. Compare FGARAT sequences to investigate the evolution of this enzyme.

- Identify the location of residues conserved between different enzyme forms.

Hypothesis: Residues conserved between different enzyme forms will be located in regions where the domain organization is shared by those forms.

Methods

All completed genomes (as of 3/1/07) in the NCBI Microbial Genomes Database were searched for FGARAT sequences, either using existing annotation or by performing BLAST searches. After initial compilation of predicted ORF sizes, the FGARAT sequences from at least one representative of each taxonomic family were retrieved from the NCBI Database, and the sequences from multiple subunit FGARATs were assembled to mimic the domain organization of the monomer. Multiple sequence alignments (Clustal W) and phylogenetic trees were generated using the Megalign component of the Lasergene sequence analysis suite (DNASTAR). The atomic coordinates for FGARAT were retrieved from the RCSB Protein Data Bank (PDB ID 1T3T - Anand et al., 2004) and analyzed using Protein Explorer (Martz, 2002)

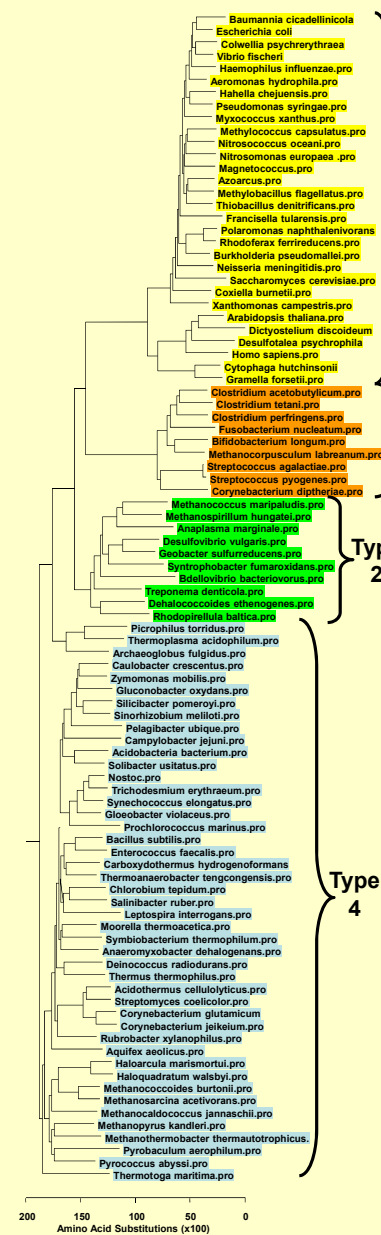


Figure 2. Phylogenetic Tree Based on Clustal W alignment of combined PurS-PurS-PurL-PurQ sequences.

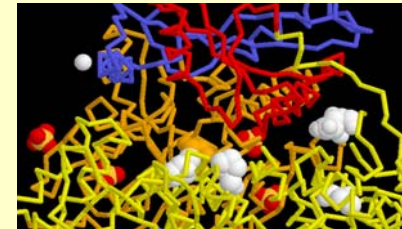


Figure 3. Residues conserved between Type 2 and Type 1b are located at the interface between PurS and PurL domains

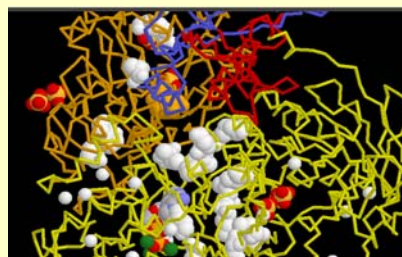


Figure 4. Residues conserved between Type 2 and Type 4 are located at the interface between PurQ and PurL domains

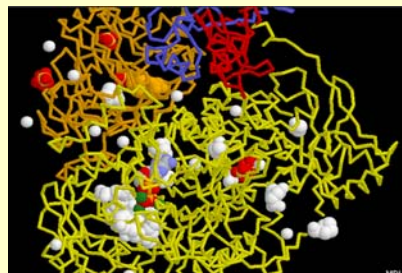


Figure 5. Residues conserved between Type 1a, 2 and Type 4 are located near the ADP binding site of the PurL domain.

Implications

The *Salmonella* PurL domain contains two active sites, one is functional, and one is vestigial but still binds ADP (Anand et al., 2004). The conservation of residues near the vestigial site in the type 1a, 2 and 4 enzymes but not in the type 1b enzyme, suggests that the type 1b enzyme may no longer bind ADP.

References

- Anand, R., Hoskins, A., Stubbe, J., Ealick, S.A. (2004). Domain Organization of *Salmonella typhimurium* Formylglycinamide Ribonucleotide Amidotransferase Revealed by X-ray Crystallography. *Biochemistry* 43:10328-10342.
- Martz, Eric. 2002. Protein Explorer: Easy Yet Powerful Macromolecular Visualization. *Trends in Biochemical Sciences*, 27:107-109.